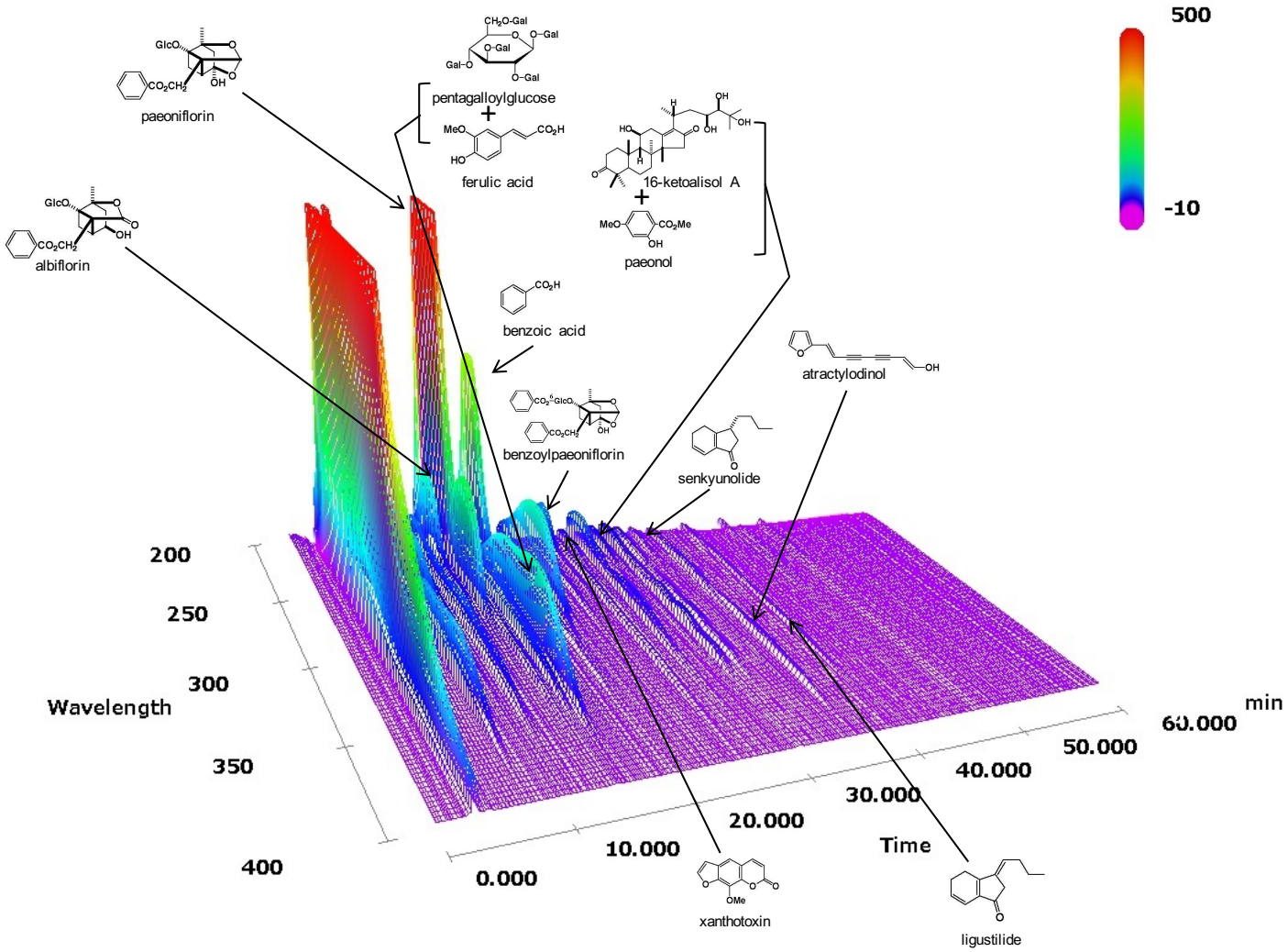
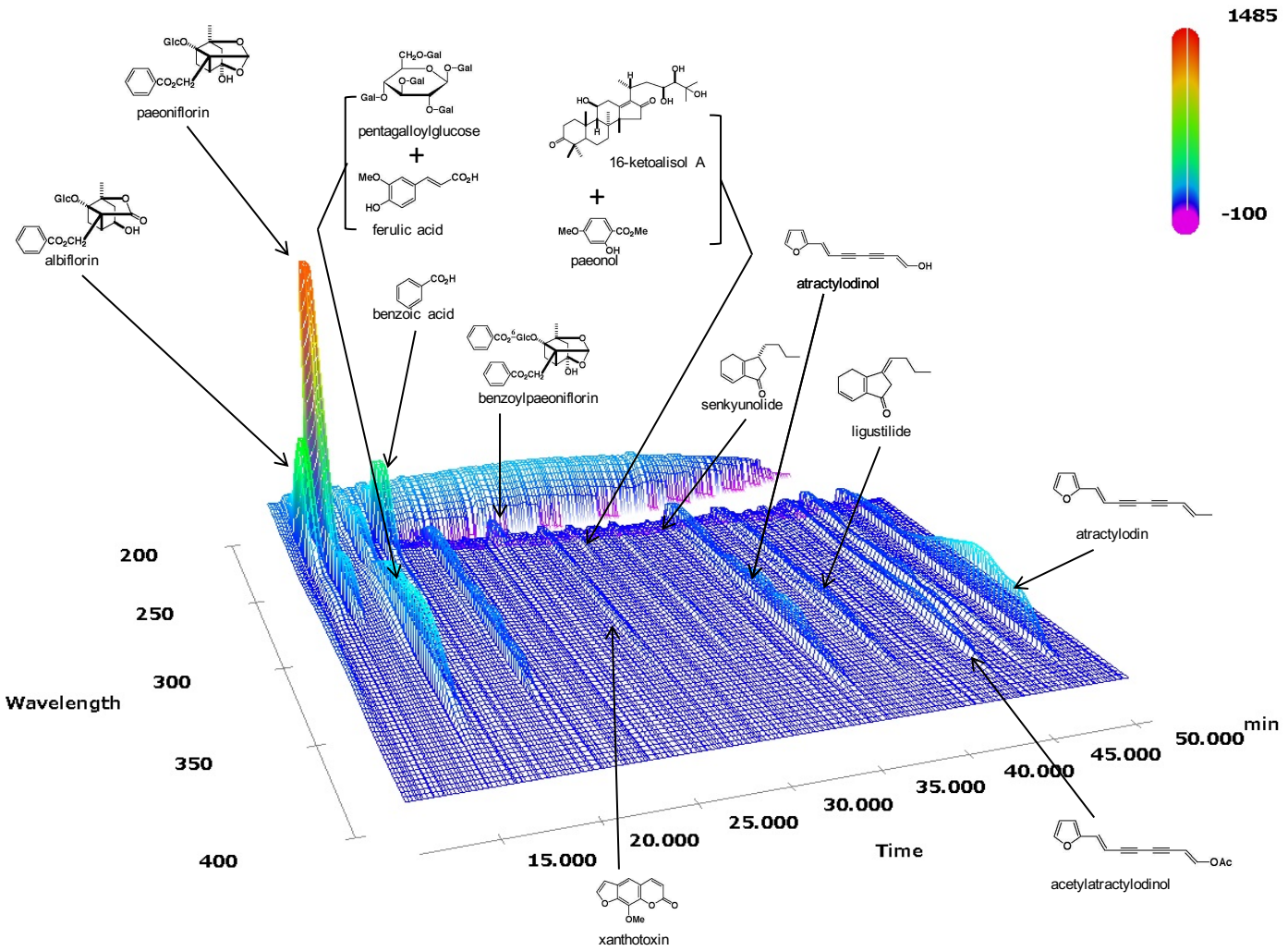
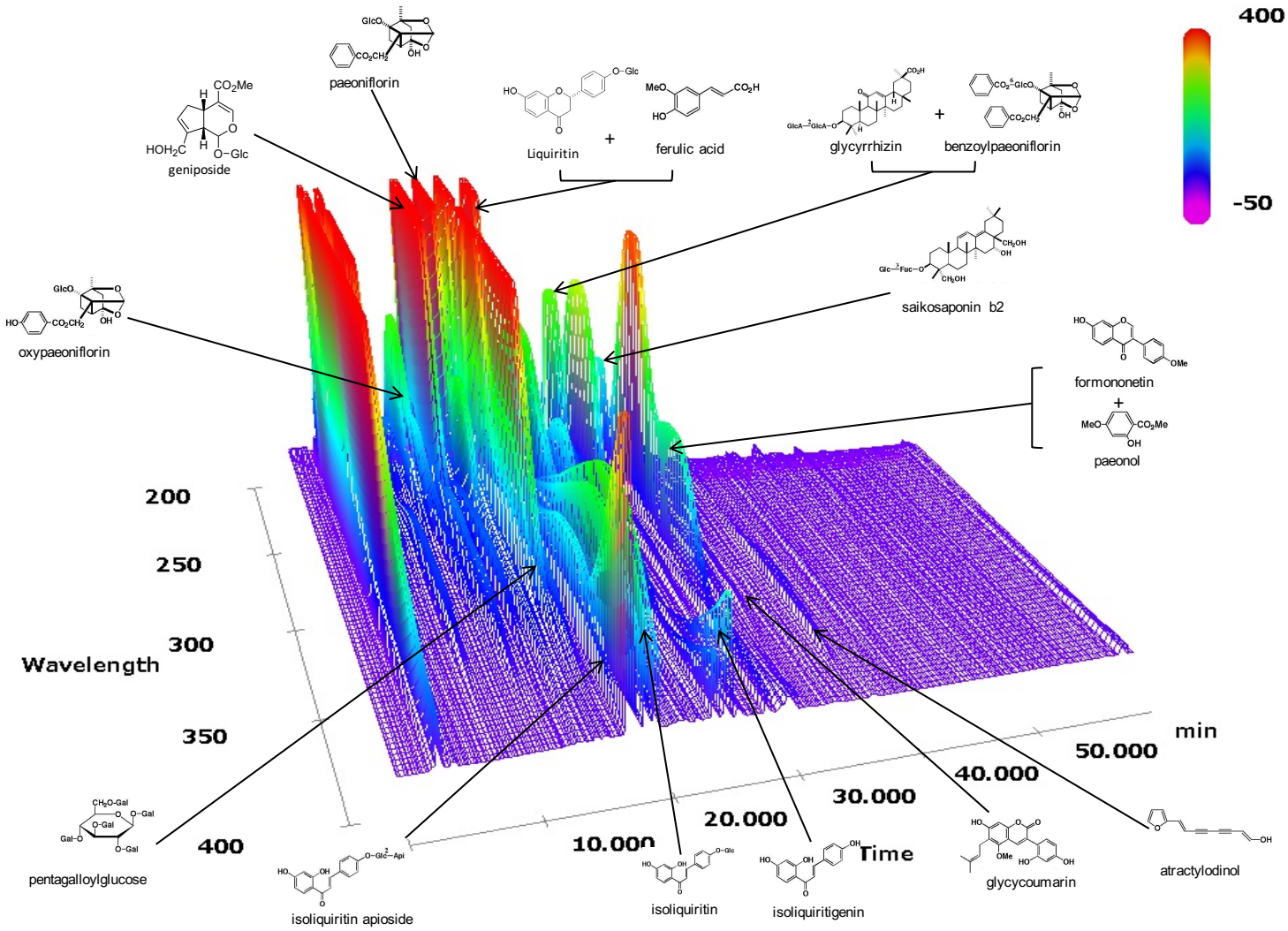


Supplemental Fig. 1
The extract of tokishakuyakusan prepared from crude drug mixture (25 mg) was suspended with MeOH (1 ml) and sonicated for 30 min. The supernatant (25 μl) was injected to HPLC with the following conditions: system, Shimadzu LC–10A$_{VP}$ (Kyoto, Japan); column, TSK–GEL ODS–80$_{TS}$ (4.6 × 250 mm, Tosoh, Tokyo); mobile phase, 0.05 M AcOH–AcONH$_4$ buffer (pH 3.6)/CH$_3$CN 90:10 (0 min) – 0:100 (60 min), linear gradient; flow rate, 1.0 ml/min; column temperature, 40°C; and detection, 200 – 400 nm by a photodiode array detector. Some peaks were identified by the retention times and UV spectra of the standard compounds.

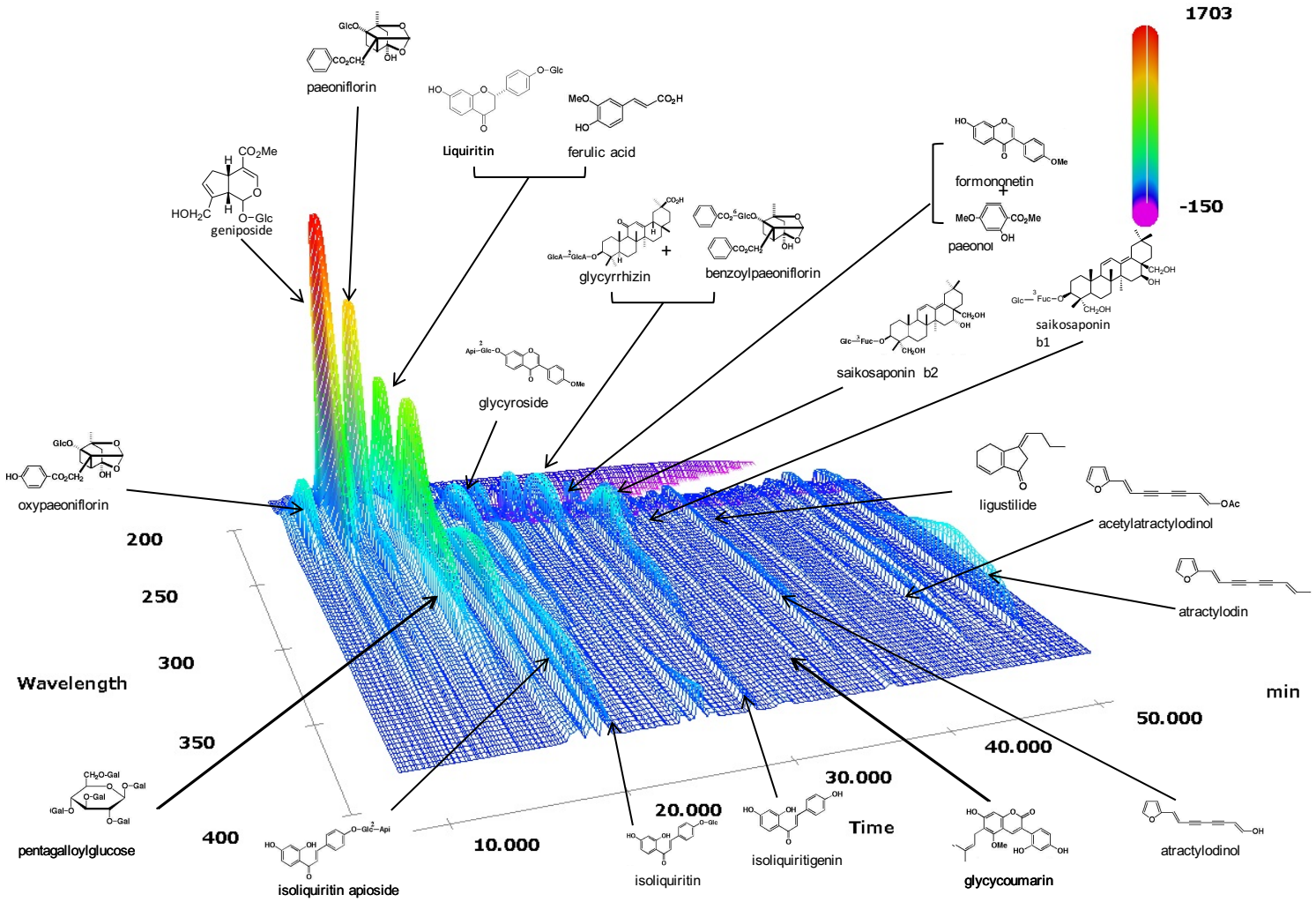


Supplemental Fig. 2
The extract of tokishakuyakusan supplied from Tsumura Co., Ltd. (25 mg) was suspended with MeOH (1 ml) and sonicated for 30 min. The supernatant (25 μl) was injected to HPLC with the following conditions: system, Shimadzu LC–10A$_{VP}$ (Kyoto, Japan); column, TSK–GEL ODS–80$_{TS}$ (4.6 × 250 mm, Tosoh, Tokyo); mobile phase, 0.05 M AcOH–AcONH$_4$ buffer (pH 3.6)/CH$_3$CN 90:10 (0 min) – 0:100 (60 min), linear gradient; flow rate, 1.0 ml/min; column temperature, 40°C; and detection, 200 – 400 nm by a photodiode array detector. Some peaks were identified by the retention times and UV spectra of the standard compounds.

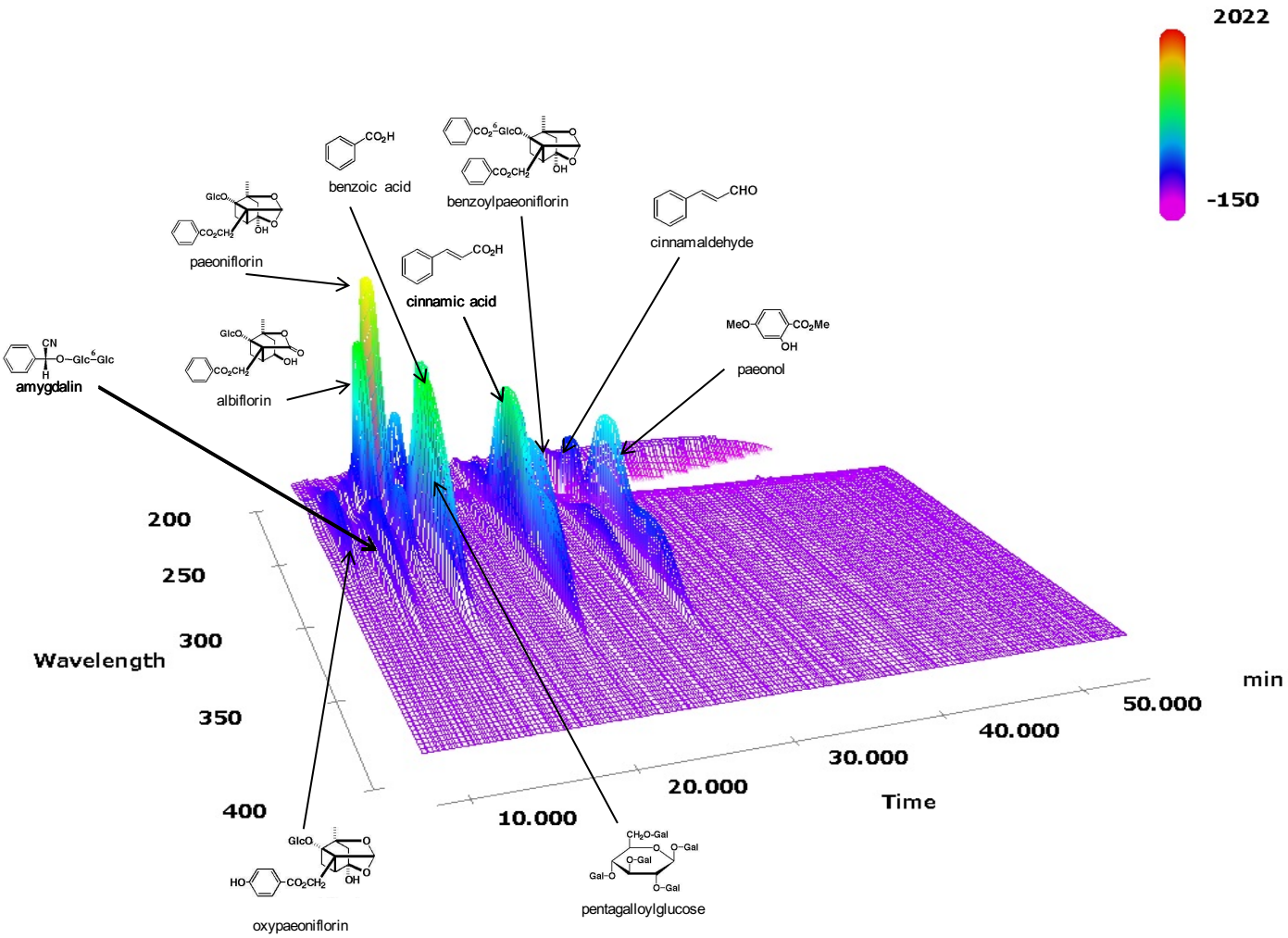


Supplemental Fig. 3

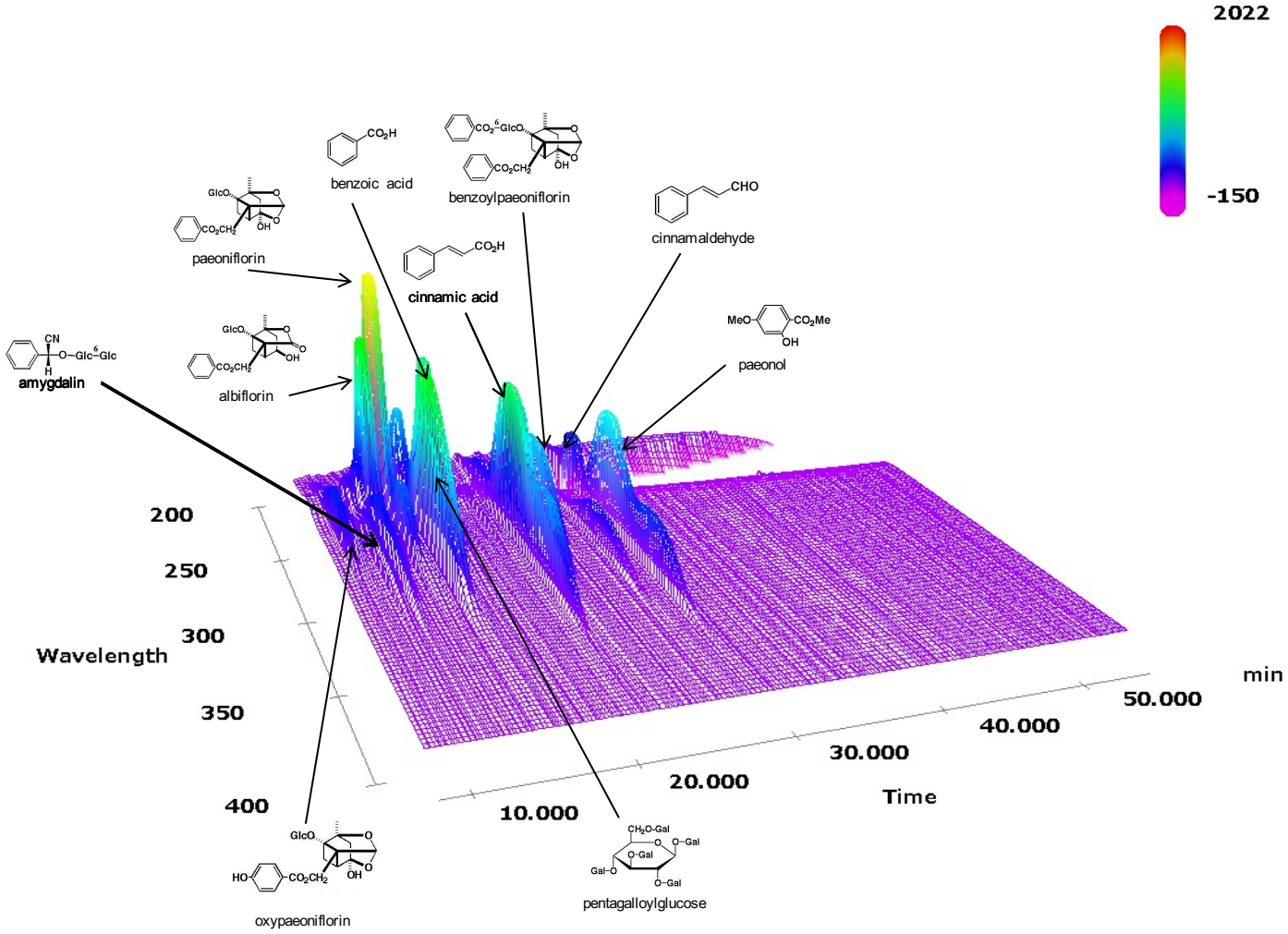
The extract of kamishoyosan prepared from crude drug mixture (25 mg) was suspended with MeOH (1 ml) and sonicated for 30 min. The supernatant (25 µl) was injected to HPLC with the following conditions: system, Shimadzu LC–10A$_{VP}$ (Kyoto, Japan); column, TSK–GEL ODS–80$_{TS}$ (4.6 × 250 mm, Tosoh, Tokyo); mobile phase, 0.05 M AcOH–AcONH$_4$ buffer (pH 3.6)/CH$_3$CN 90:10 (0 min) – 0:100 (60 min), linear gradient; flow rate, 1.0 ml/min; column temperature, 40°C; and detection, 200 – 400 nm by a photodiode array detector. Some peaks were identified by the retention times and UV spectra of the standard compounds.



Supplemental Fig. 4
The extract of kamishoyosan supplied from Tsumura Co., Ltd. (25 mg) was suspended with MeOH (1 ml) and sonicated for 30 min. The supernatant (25 µl) was injected to HPLC with the following conditions: system, Shimadzu LC–10A_{VP} (Kyoto, Japan); column, TSK–GEL ODS–80$_{TS}$ (4.6 × 250 mm, Tosoh, Tokyo); mobile phase, 0.05 M AcOH–$AcONH_4$ buffer (pH 3.6)/CH_3CN 90:10 (0 min) – 0:100 (60 min), linear gradient; flow rate, 1.0 ml/min; column temperature, 40°C; and detection, 200 – 400 nm by a photodiode array detector. Some peaks were identified by the retention times and UV spectra of the standard compounds.



Supplemental Fig. 5
The extract of keishibukuryogan prepared from crude drug mixture (25 mg) was suspended with MeOH (1 ml) and sonicated for 30 min. The supernatant (25 µl) was injected to HPLC with the following conditions: system, Shimadzu LC–10A$_{VP}$ (Kyoto, Japan); column, TSK–GEL ODS–80$_{TS}$ (4.6 × 250 mm, Tosoh, Tokyo); mobile phase, 0.05 M AcOH–AcONH$_4$ buffer (pH 3.6)/CH$_3$CN 90:10 (0 min) – 0:100 (60 min), linear gradient; flow rate, 1.0 ml/min; column temperature, 40°C; and detection, 200 – 400 nm by a photodiode array detector. Some peaks were identified by the retention times and UV spectra of the standard compounds.



Supplemental Fig. 6
The extract of keishibukuryogan supplied from Tsumura Co., Ltd. (25 mg) was suspended with MeOH (1 ml) and sonicated for 30 min. The supernatant (25 µl) was injected to HPLC with the following conditions: system, Shimadzu LC–10A$_{VP}$ (Kyoto, Japan); column, TSK–GEL ODS–80$_{TS}$ (4.6 × 250 mm, Tosoh, Tokyo); mobile phase, 0.05 M AcOH–AcONH$_4$ buffer (pH 3.6)/CH$_3$CN 90:10 (0 min) – 0:100 (60 min), linear gradient; flow rate, 1.0 ml/min; column temperature, 40°C; and detection, 200 – 400 nm by a photodiode array detector. Some peaks were identified by the retention times and UV spectra of the standard compounds.